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## **Nucleosides, Nucleotides and Nucleic Acids**

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## **Polyfunctional O-Substituted Hydroxylamines: Modification of Nucleic Acids, Inhibition of Sam-Decarboxylase**

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POLYFUNCTIONAL O-SUBSTITUTED HYDROXYLAMINES: MODIFICATION  
OF NUCLEIC ACIDS, INHIBITION OF SAM-DECARBOXYLASE

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ABSTRACT. Treo-1-aminooxy-2,3-dihydroxy-4-mercaptobutane is suggested for the introduction of reactive thiol groups via cytidine residues and/or 3'-end of nucleic acids. S-(5'-desoxyadenosyl)-aminooxyethyl-thiomethyl hydroxylamine irreversibly inhibits SAM decarboxylase in  $10^{-5}$  M.

Many reactions for modification of nucleic acids were suggested, but approaches for base-specific introduction of reactive groups (HS-;  $H_2N$ -;  $HOOC$ -; etc.) were not developed. Among the functions to be introduced into nucleic acids the most promising are HS- and  $H_2N$ -groups, because of high reactivity under mild conditions and sensitivity of quantitative determinations. Hydroxylamine and O-methylhydroxylamine are classic reagents for RNA and DNA modification with established mechanism of reaction through cytidine residues. So, we constructed a series of polyfunctional hydroxylamines  $H_2N-O-R-X$  ( $X = -SH$ ;  $-NH_2$ ;  $-COOH$ ; etc.) that reacted with tRNA like O-methylhydroxylamine.

O-methylhydroxylamine modifies DNA and RNA only at high concentrations, so hydroxylamines  $H_2N-O-R-X$  must be good soluble at neutral and slightly acidic pH (optimum is 5.0) remaining uncharged. Treo-1-aminooxy-2,3-dihydroxy-4-mercaptobutane (aminooxythiotreitol) /1/ is the simplest structure, being in accordance with the above requirements and it was used for nucleic acids modification.

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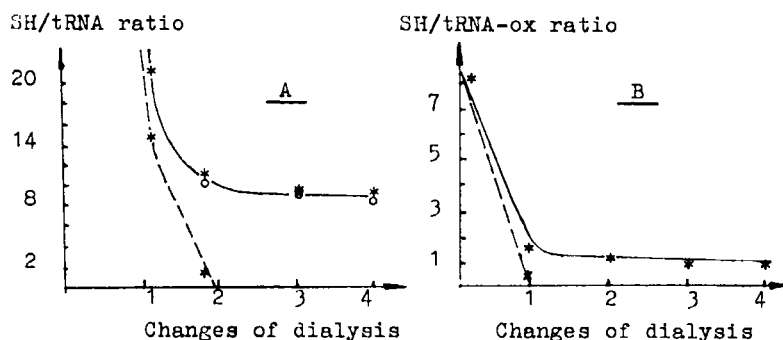


FIG. 1. Introduction of HS-groups into tRNA cytidine residues (A) and 3'-oxidised ribose (B).  
 A: 0,82 M aminooxythiotreitol, pH 5,0;  $2,5 \cdot 10^{-4}$  M tRNA (165 o.u./ml) were incubated 16 hr. at  $37^\circ$  (under these conditions 14-C-CH<sub>3</sub>ONH<sub>2</sub>:tRNA ratio is 10:1) and dialysed against 1 mM EDTA and 50 mM NaCl for 48 hr. with four buffer changes. SH/tRNA ratio was determined with Ellman reagent (\*) and o-chloromercuri-p-nitrophenol (o). In the control the same mixture was dialysed on being prepared.  
 B:  $5,0 \cdot 10^{-4}$  M tRNA-ox (330 o.u./ml) and  $3,5 \cdot 10^{-3}$  M 1-amino-oxy-4-mercaptobutane in 0,1 M Na-acetate pH 5,0 were incubated 2 hr. at  $20^\circ$ . Determination of SH/tRNA ratio is described in A. In control tRNA was used instead of tRNA-ox.

Incubation of tRNA in concentrated aminooxythiotreitol solutions resulted in incorporation of eight reactive thiol groups per mole of tRNA /2/, being titrated with Ellman reagent and o-chloromercuri-p-nitrophenol (Fig. 1A)

1-aminooxy-4-aminobutane reacted with tRNA by the same way and introduced amino groups were determined by fluorescamine titration (Fig. 2).

Polyfunctional O-substituted hydroxylamines turned out to be convenient reagents for introduction of reactive groups via 3'-oxidised terminal of RNA. The morpholidate-type adducts with dilute solutions of 14-C-CH<sub>3</sub>ONH<sub>2</sub> were formed rapidly and specifically through 3'-end. We have introduced via 3'-terminal reactive: HS- and H<sub>2</sub>N-groups (Fig. 1B and Fig. 2), as well as HOOC-; (HO)<sub>2</sub>(O)P-; H<sub>2</sub>NO-groups. <sup>125</sup>I-label and polymercurated cluster were introduced by means of earlier unknown mercury-containing hydroxylamines /3/.

One of the key enzymes in polyamine biosynthesis is pyruvate-dependent decarboxylase of S-adenosylmethionine

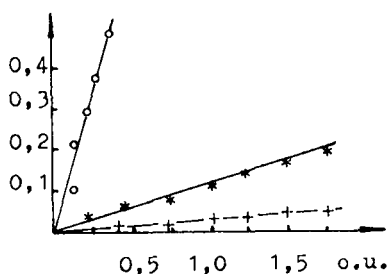


FIG. 2.

Introduction of  $H_2N$ -groups into tRNA.  
 $1,2 \cdot 10^{-4}$  M tRNA and tRNA-ox were incubated at pH 5,0 in 1,0 M and  $1,2 \cdot 10^{-3}$  M 1-aminooxy-4-aminobutane as described in Fig.1(A,B). After the removal of the excess of the reagent  $H_2N$ -groups introduced in cytidine residues (o) or 3'-oxydised ribose (\*) were titrated with fluram ( $\lambda_{ex}$  385;  $\lambda_{em}$  485). (+) is the control.

TABLE 1. Irreversible inhibition of SAM decarboxylase from E.Coli and rat liver.

Compound	Preincubation 30 min ( $I_{50}$ )	
	E.Coli	Rat liver
Ado-S-( $CH_2$ ) <sub>2</sub> -ONH <sub>2</sub> (I)   CH <sub>3</sub>	$2 \cdot 10^{-8}$ M	$3 \cdot 10^{-9}$ M
Ado-S-( $CH_2$ ) <sub>2</sub> -ONH <sub>2</sub> (II)	$1 \cdot 10^{-4}$ M	$4 \cdot 10^{-5}$ M
Ado-S-( $CH_2$ ) <sub>4</sub> -ONH <sub>2</sub> (III)   CH <sub>3</sub>	$9 \cdot 10^{-7}$ M	$8 \cdot 10^{-9}$ M
Ado-S-( $CH_2$ ) <sub>4</sub> -ONH <sub>2</sub> (IV)	$5 \cdot 10^{-4}$ M	$2 \cdot 10^{-4}$ M
H <sub>3</sub> C-S-( $CH_2$ ) <sub>2</sub> -ONH <sub>2</sub> (V)   CH <sub>3</sub>	$2 \cdot 10^{-4}$ M	-----
H <sub>3</sub> C-S-( $CH_2$ ) <sub>2</sub> -ONH <sub>2</sub> (VI)	$5 \cdot 10^{-4}$ M	-----

(SAM). Hydroxylamine-containing analogs of decarboxylated SAM were synthesised /1/ as a means for irreversible inhibition of this enzyme /4,5/.

Substance (I) was found to be 1000 times more active than the best among known inhibitors of this enzyme. The inhibition was time-dependent, substrate was incapable to restore the enzyme activity, but it protected enzyme from (I) action. Application of Kitz-Willson's approximation for the determination of the inhibition parameters indicated on double step process. The affinity of compound (I) towards enzyme active site at the integral reversible step was 100 times higher than decarboxylated SAM had. Substance (I) interacts with the enzyme in monocation form ( $pK_{H_2NO}$ -group is 4,5-5,0) that seems to be preferable in comparison with double cation form of decarboxylated SAM.

Specific mode of SAM-decarboxylase interaction with (I) is confirmed by low activities of (II) and (V) towards enzyme. Besides, pyridoxal-5'-phosphate dependent aspartate aminotransferase is slightly inhibited at mM concentrations of compound (I).

High potency and specificity of (I) towards SAM decarboxylase can be explained in terms of similarity between E-I complex and one of E-S intermediates.

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